



Recent developments and innovations in red blood cells diagnostics

Giuseppe Lippi¹, Mario Plebani²

¹Section of Clinical Biochemistry, University of Verona, Verona, Italy; ²Department of Laboratory Medicine, University Hospital of Padova, Padova, Italy

Correspondence to: Giuseppe Lippi, MD. Section of Clinical Biochemistry, University Hospital of Verona, Piazzale LA Scuro 10, Verona 37134, Italy. Email: giuseppe.lippi@univr.it.

Abstract: Although the diagnosis of anemia is a relatively simple and straightforward endeavor, since it is mostly based on the measurement of hemoglobin in whole blood, the accurate characterization of red blood cell (RBC) disorders has emerged as a mainstay in the era of precision (laboratory) medicine. The many technological advances occurred in laboratory medicine over recent times have enabled the introduction of a vast array of innovations also for RBC diagnostics, mostly represented by modular automation, digital hematology, innovative erythrocyte parameters and adaptation of disruptive technologies to laboratory hematology, especially mass spectrometry and molecular biology. These important breakthroughs have enormously contributed to broadening the diagnostic armamentarium and making more efficient and sustainable the diagnostics of RBC disorders. Some on these important innovations will be discussed in this article.

Keywords: Hemoglobin; red blood cells (RBCs); anemia; diagnosis; laboratory

Received: 19 July 2018; Accepted: 30 July 2018; Published: 10 August 2018.

doi: 10.21037/jlpm.2018.07.09

View this article at: <http://dx.doi.org/10.21037/jlpm.2018.07.09>

Introduction

Anemia, which is currently defined by the World Health Organization (WHO) as a hemoglobin value in whole blood <120 g/L in women and <130 g/L in men, respectively (1), can now be regarded as a worldwide endemic disease, with an estimated prevalence of 1.62 billion people, which approximates 25% of the worldwide population (2). Albeit iron deficiency is the leading cause of anemia (i.e., approximating 50% of cases), this condition is rarely present alone, but may coexist with a kaleidoscope of other causes such as nutritional deficiencies (i.e., folate or vitamin B12 deficiencies), acute or chronic bleeding, hereditary red blood cell (RBC) disorders (i.e., hemoglobinopathies, spherocytosis), infections, renal or liver impairment, cancer and chronic inflammatory conditions (*Table 1*) (1).

Since the clinical signs and symptoms of anemia are often poorly specific, and may also be subtle, especially

in patients with chronic forms of anemia, laboratory hematology represents a virtually unavoidable part of both the diagnostic reasoning and clinical decision making, since laboratory tests provide irreplaceable information for screening, diagnosis and monitoring of RBC disorders (3,4). Beside the conventional measurement of whole blood hemoglobin content, which is a necessary precondition for diagnosing anemia, the preliminary classification of anemias is usually based on the values of mean corpuscular volume (MCV) and RBC distribution width (RDW) (5), as summarized in *Table 1*. This approach is still forthright and valid, but does not enable a definitive etiological characterization, and should hence be complemented with a panel of additional laboratory investigations. Importantly, a number of technological advances occurred over the past few decades have enormously contributed to broadening the diagnostic armamentarium and making more efficient and sustainable the diagnostics of RBC disorders. Some on these important innovations will be summarized in the following

Table 1 Classification of anemia according to RDW and MCV

Condition	RDW	MCV
Nutritional deficiencies		
Iron deficiency	↑	↓
Folic acid deficiency	↑	↑
Vitamin B deficiency	↑	↑
β-thalassemia	↑	↓
Hemolytic anemias		
Immune hemolytic anemia	↑	↑
Hereditary spherocytosis	N/↑	N/↓
Anemic hemoglobinopathies (i.e., SS, SC)	↑	N
Sickle cell trait	N	↓
Chronic disorders		
Chronic diseases anemia	N	↓
Chronic liver disease	↑	N/↑
Hematologic disorders		
Aplastic anemia	N	↑
Chronic leukemias	N	N
Myelodysplastic syndrome	↑	↑
Other thalassemias	N	↓
Acute hemorrhages	N	N

↑, increased; N, normal; ↓, decreased. RDW, red blood cell distribution width; MCV, mean corpuscular volume.

parts of this article.

Automation of laboratory hematology

Laboratory automation should be regarded as one of the major advancements occurred in RBC diagnostics. Laboratory automation is conventionally defined as a multi-disciplinary integration of robotics, information technology (IT), sample handling and many other technologies aimed at developing and optimizing both workflow and activities within medical laboratories (6). Briefly, laboratory automation may bring paramount benefits to *in vitro* diagnostic testing, including easier and more efficient management of workflows, better withstanding the increasing complexity and volume of routine and urgent testing, improved turnaround time (TAT), dismissal of many manual activities, enhanced walk-away, cost savings (i.e., especially those attributable to subsidiary staff and

technicians), improved standardization of procedures, lower chance of errors throughout the total testing process, decreased biological risk, along with opportunities to implement automatic reruns or reflex testing (7).

Laboratory automation has been for long limited to clinical chemistry and immunochemistry platforms. The major hurdle encountered in developing models of automation for laboratory hematology is represented by the peculiar sample type, which is whole blood anticoagulated with dipotassium ethylenediaminetetraacetic acid (EDTA) (8). The presence of EDTA in the sample, which irreversibly sequesters ionized calcium and many other metal ions (9), makes EDTA plasma an unsuitable sample matrix for clinical chemistry and even for coagulation testing, thus generating considerable obstacles for consolidation of laboratory hematology with other branches of laboratory medicine. Nevertheless, a number of technological solutions have been developed for laboratory hematology in recent times. These basically include (I) commercialization of modular, high-throughput and versatile analyzers, which can be easily interconnected by means of sample conveyers, and can fit the organization of small, medium and large facilities, (II) integration of preanalytical workstations, which can be identical to those included in models of total laboratory automation, or can be specifically designed to suit hematological testing, (III) connection with automated slide strainers, which help improving the entire slide making process (i.e., less manual activities and lower biological risk, improved standardization of slide preparation and staining, customization of staining protocols, reduction of TAT) (10), as well as (IV) integration of automated image analysis systems (see the following section of this article). This organization is often referred to as “modular laboratory hematology” (Figure 1).

Digital hematology

Throughout the relatively long history of laboratory hematology, the only reliable means for identifying, enumerating and sizing blood cells has been for long represented by optical microscopy of peripheral blood smear. This practice carries many drawbacks, since it is inevitably time-consuming, is vulnerable to high inter- and intra-observer inaccuracy and imprecision, needs specific education and training of microscopists, and is poorly suited for rapid diagnostics as otherwise needed in patients with many acute hematological disorders (11).

Recent technological advances have led to development

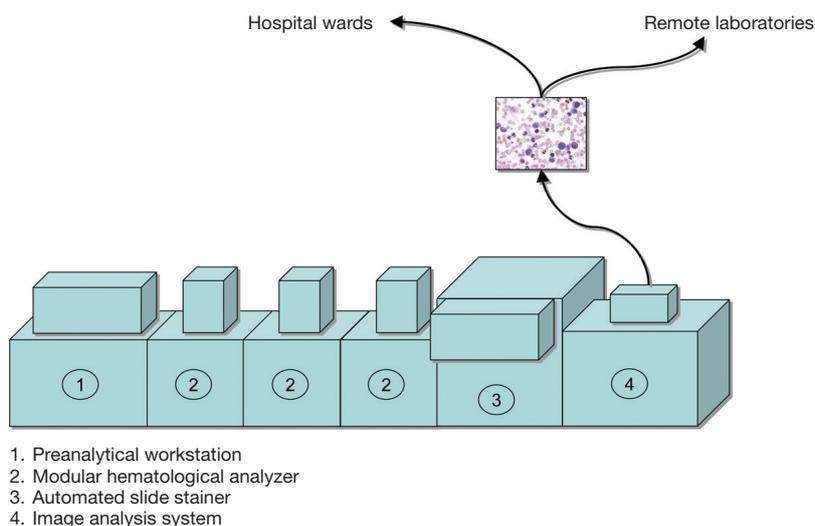


Figure 1 Modular laboratory hematology.

and commercialization of innovative automated image analysis systems, which are suited for automation and can hence be directly connected (in series) with hematologic analyzers (*Figure 1*) (12). These innovative platforms scan the slides (usually at a picture of $\times 100$ objective), and store digitalized images of blood smears at high magnification. The images are analyzed by artificial neural networks based on a preexisting database of blood elements (thus including RBC), which can be locally customized or updated by the users. The images can be transmitted to, and displayed on, computer screens, which can be even placed at long distances from the scanner (i.e., in hospital wards or in remote laboratories) (*Figure 1*), for analysis and potential reclassification of blood elements. The operator can also increase the size of the images, or expand single sections of the scan, so obtaining a more accurate view. The operator can then accept and conserve the automatic classification, or can move elements from one cell category to another, thus improving the final reclassification. Albeit these automated image analysis systems have been originally developed for analysis of white blood cells (WBC), specific information can also be garnered on erythrocyte morphology, thus including the presence of anisocytosis, hypochromia, microcytosis or macrocytosis, spherocytosis, elliptocytosis, ovalocytosis, stomatocytosis, acanthocytosis, echinocytosis, polychromasia, poikilocytosis and abnormal erythrocytes (i.e., sickle cells and schizocytes, helmet and teardrop cells) (13). Recent data showed that the diagnostic sensitivity of these systems for identifying some critical

categories of abnormal erythrocytes (i.e., spherocytes or sickle, target and tear drop cells) is excellent, typically higher than 80% (14), thus making the use of digital image analysis a highly valuable, and probably more accurate and reproducible, alternative to optical microscopy.

Notably, the use of these systems may also enable an efficient recognition of parasitoid infections such as Malaria (15), as well as the reliable identification of intravascular and spurious hemolysis, which would be otherwise undetectable on whole blood specimens (16,17). Interestingly, most of these automated image analysis systems are also capable of optimizing the identification of rare RBC abnormalities, since morphological erythrocyte alterations can be more efficiently visualized on the computer screen (18). Finally, the creation of a large personalized database of images of suggestive RBC abnormalities represents a valuable resource for education and training of students and laboratory professionals (19).

Innovative erythrocyte parameters

Irrespective of the fact that the diagnosis of anemia is relatively simple and straightforward (by measuring total hemoglobin in whole blood), the newer generation of hematologic analyzers is now equipped with many analytical and technical innovations, which enable obtaining other information than that reported with the traditional complete blood cell count (CBC), and which may ultimately provide a substantial improvement for the differential diagnosis

of anemias (20). Although a more detailed discussion will be omitted for space constraints, some important aspects deserve a special mention. These innovative parameters most typically include automated reticulocyte and nucleated RBC counts, hemoglobinization of reticulocytes and RBC, reticulocyte hemoglobin content (occasionally defined as CHr and RET-He according to the technology used for its assessment), reticulocyte maturation, automatic analysis and calculation of microcytic and hypochromic RBC (21,22). The various combination of these different parameters not only may be useful to complement clinical history, physical examination and results of more conventional laboratory investigations (i.e., CBC, ferritin, transferrin, iron, haptoglobin, folic acid and vitamin B12, among others) for troubleshooting the underlying cause(s) of anemia (23), but may also be clinically useful for diagnosing, prognosticating and monitoring other non-RBC disorders, as recently shown for patients with sepsis (24), chronic kidney disease (25) and cancer (26). The generation of complex scattergrams (27,28), which is now almost commonplace in the vast majority of hematologic analyzers, is also helpful for more accurately identifying abnormal RBC populations and other atypical elements, as recently shown for diagnosing malaria (29).

Disruptive technologies

Regardless of consolidated laboratory techniques, which have just recently made their way through phenotypic diagnostics of anemia (i.e., capillary electrophoresis) (30), a major innovation has been represented by the application of mass spectrometry and molecular biology in the diagnostics of hemoglobinopathies. The former approach allows a better characterization of hemoglobin variants preliminarily identified by screening techniques such as high-pressure liquid chromatography (HPLC) or capillary electrophoresis (31), whilst molecular diagnostic techniques enable to unravel specific molecular abnormalities characterizing many congenital RBC disorders (32).

Unlike screening tests, the selection of the most appropriate molecular diagnostic approach in patients with inherited hemoglobinopathies or RBC enzymopathies should take into account the prevalence and penetrance of the different mutations in the ethnic populations, across different geographical locations. Therefore, the first step may be represented by polymerase chain reaction (PCR)-based techniques [e.g., restriction-endonuclease PCR

(RE-PCR), amplification refractory mutation system (ARMS), resolution melting analysis (HRMA), denaturing gradient gel electrophoresis (DGGE)]. Allele-specific methodologies, such as allele-specific PCR and reverse dot-blot, are especially useful for thalassemia diagnostics in target populations, enable processing a high volume of samples and are relatively inexpensive, permitting to screen some prevalent hemoglobin genes mutations at the same time. Array comparative genomic hybridization (array CGH) can then be used for detecting additional mutations which cannot be identified with first-line DNA analysis. Regarding thalassemias, gap-PCR (gap-PCR) and multiplex ligation-dependent probe amplification (MLPA) are perhaps the best options for screening and also for detecting large deletions or duplications of globin genes, which cannot be identified with conventional DNA sequencing. Sanger or next-generation sequencing (NGS) techniques may then be particularly suited for detecting all known point-mutations, but may also enable indentifying novel or rare mutations, thus helping to uncover new mechanisms of disease. A reliable guidance for the cost-effective integration of these different molecular techniques has recently been published by the European Molecular Genetics Quality Network (EMQN) (33). Notably, emerging evidence also suggests that molecular genetic testing has a pivotal role in patients with diseases characterized by clonal hematopoiesis, thus supporting the diagnostic workout of hematologic malignancies and/or myelodysplastic syndromes (34).

Conclusions

Albeit the laboratory diagnostics of anemia remains a rather simple enterprise, accurate disease characterization has emerged as a mainstay in the era of precision (laboratory) medicine, even for RBC disorders (35). The many technological advances occurred in laboratory medicine over recent times have enabled the introduction of a vast array of innovations (*Table 2*), which have led the way to a more efficient patient care and a more convenient organization of resources and workflows within the laboratory. In the foreseeable future, the better understanding of phenotypic heterogeneity of RBC disorders, also supported by IT tools such as expert diagnostic systems (36) or artificial neural networks (37), will predictably enable to improve the global management of these disorders at multiple levels. Yet, some additional issues will need to be addressed, on top of it all the current lack of harmonization in laboratory hematology (11).

Table 2 Recent developments in RBCs diagnostics

Automation
Modular, high-throughput analyzers
Integration of preanalytical workstations
Connection with automated slide strainers
Integration of automated image analysis systems
Digital image analysis
High diagnostic sensitivity
Improved identification of rare erythrocyte abnormalities
Enhanced recognition of parasitoid infections
Reliable identification of intravascular and spurious hemolysis
Possible use of the digitalized database for education and training
Innovative erythrocyte parameters
Automated reticulocyte count
Nucleated red blood cells count
Hemoglobinization of erythrocytes and reticulocytes
Reticulocyte hemoglobin content
Reticulocyte maturation
Microcytic and hypochromic erythrocytes
Low hemoglobin density
Advanced scattergrams
Disruptive technologies
Capillary electrophoresis
Mass spectrometry
Molecular diagnostics
RE-PCR
ARMS
HRMA
DGGE
Allele-specific PCR and reverse dot-blot
Array CGH
Gap-PCR
MLPA
Sanger or NGS
IT
Expert diagnostic systems
Artificial neural networks

RBC, red blood cell; RE-PCR, restriction-endonuclease polymerase chain reaction; ARMS, amplification refractory mutation system; HRMA, resolution melting analysis; DGGE, denaturing gradient gel electrophoresis; CGH, comparative genomic hybridization; MLPA, multiplex ligation-dependent probe amplification; NGS, next-generation sequencing; IT, information technology.

Acknowledgments

Funding: None.

Footnote

Provenance and Peer Review: This article was commissioned by the editorial office, *Journal of Laboratory and Precision Medicine* for the series “Laboratory Medicine-25 Years on”. The article has undergone external peer review.

Conflicts of Interest: Both authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/jlpm.2018.07.09>). The series “Laboratory Medicine-25 Years on” was commissioned by the editorial office without any funding or sponsorship. Giuseppe Lippi served as an unpaid Guest Editor of the series and serves as the unpaid Editor-in-Chief of *Journal of Laboratory and Precision Medicine* from November 2016 to October 2021. Mario Plebani served as an unpaid Guest Editor of the series. The authors have no other conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

1. World Health Organization. Haemoglobin concentrations for the diagnosis of anaemia and assessment of severity. Vitamin and Mineral Nutrition Information System. World Health Organization, Geneva, Switzerland, 2011.
2. de Benoist B, McLean E, Egli I, et al. Worldwide prevalence of anaemia 1993-2005. In: WHO Global Database on Anaemia. World Health Organization, Geneva, Switzerland, 2008.

3. Coyer SM. Anemia: diagnosis and management. *J Pediatr Health Care* 2005;19:380-5.
4. Guidi GC, Lechi Santonastaso C. Advancements in anemias related to chronic conditions. *Clin Chem Lab Med* 2010;48:1217-26.
5. Bessman JD, Gilmer PR Jr, Gardner FH. Improved classification of anemias by MCV and RDW. *Am J Clin Pathol* 1983;80:322-6.
6. Olsen K. The first 110 years of laboratory automation: technologies, applications, and the creative scientist. *J Lab Autom* 2012;17:469-80.
7. Genzen JR, Burnham CD, Felder RA, et al. Challenges and Opportunities in Implementing Total Laboratory Automation. *Clin Chem* 2018;64:259-64.
8. International Council for Standardization in Haematology. Recommendations of the International Council for Standardization in Haematology for Ethylenediaminetetraacetic Acid Anticoagulation of Blood for Blood Cell Counting and Sizing. International Council for Standardization in Haematology: Expert Panel on Cytometry. *Am J Clin Pathol* 1993;100:371-2.
9. Banfi G, Salvagno GL, Lippi G. The role of ethylenediamine tetraacetic acid (EDTA) as in vitro anticoagulant for diagnostic purposes. *Clin Chem Lab Med* 2007;45:565-76.
10. de Bitencourt ED, Voegeli CF, Onzi Gdos S, et al. Validation of the Sysmex sp-1000i automated slide preparer-stainer in a clinical laboratory. *Rev Bras Hematol Hemoter* 2013;35:404-8.
11. Buoro S, Lippi G. Harmonization of laboratory hematology: a long and winding journey. *Clin Chem Lab Med* 2018. [Epub ahead of print].
12. Da Costa L. Digital image analysis of blood cells. *Clin Lab Med* 2015;35:105-22.
13. Nakashima MO, Doyle TJ, Phelan-Lewin K, et al. Assessment of semi-quantitative grading of red blood cell abnormalities utilizing images from the CellaVision DM96 compared to manual light microscopy. *Int J Lab Hematol* 2017;39:e110-2.
14. Criel M, Godefroid M, Deckers B, et al. Evaluation of the Red Blood Cell Advanced Software Application on the CellaVision DM96. *Int J Lab Hematol* 2016;38:366-74.
15. Florin L, Maelegheer K, Muyldermans A, et al. Evaluation of the CellaVision DM96 advanced RBC application for screening and follow-up of malaria infection. *Diagn Microbiol Infect Dis* 2018;90:253-56.
16. Lippi G, Pavesi F, Benegiamo A, et al. What do hemolyzed whole-blood specimens look like? Analysis with a CellaVision DM96 automated image analysis system. *J Lab Autom* 2015;20:60-3.
17. Huisjes R, van Solinge WW, Levin MD, et al. Digital microscopy as a screening tool for the diagnosis of hereditary hemolytic anemia. *Int J Lab Hematol* 2018;40:159-68.
18. Horn CL, Mansoor A, Wood B, et al. Performance of the CellaVision(®) DM96 system for detecting red blood cell morphologic abnormalities. *J Pathol Inform* 2015;6:11.
19. Shlebak AA, Bain BJ. Training future haematologists, a privilege or a burden? "A trainer's view". *Br J Haematol* 2017;178:501-7.
20. Buttarello M, Plebani M. Automated blood cell counts: state of the art. *Am J Clin Pathol* 2008;130:104-16.
21. Urrechaga E, Borque L, Escanero JF. Biomarkers of hypochromia: the contemporary assessment of iron status and erythropoiesis. *Biomed Res Int* 2013;2013:603786.
22. Schoorl M. Innovative haematological parameters in clinical practice. *Ned Tijdschr Klin Chem Labgeneesk* 2016;41:6-16.
23. Schoorl M, Schoorl M, van Pelt J, et al. Application of Innovative Hemocytometric Parameters and Algorithms for Improvement of Microcytic Anemia Discrimination. *Hematol Rep* 2015;7:5843.
24. Buoro S, Manenti B, Seghezzi M, et al. Innovative haematological parameters for early diagnosis of sepsis in adult patients admitted in intensive care unit. *J Clin Pathol* 2018;71:330-5.
25. Garzia M, Di Mario A, Ferraro E, et al. Reticulocyte Hemoglobin Equivalent: an indicator of reduced iron availability in chronic kidney diseases during erythropoietin therapy. *Lab Haematol* 2007;13:6-11.
26. Peerschke E, Pessin MS, Maslak P. Using the hemoglobin content of reticulocytes (RET-He) to evaluate anemia in patients with cancer. *Am J Clin Pathol* 2014;142:506-12.
27. Kakkar N, Makkar M. Red Cell Cytograms Generated by an ADVIA 120 Automated Hematology Analyzer: Characteristic Patterns in Common Hematological Conditions. *Lab Med* 2009;40:549-55.
28. Huh J, Moon H, Chung W. Erroneously elevated immature reticulocyte counts in leukemic patients determined using a Sysmex XE-2100 hematology analyzer. *Ann Hematol* 2007;86:759-62.
29. Buoro S, Manenti B, Seghezzi M, et al. Abnormal scattergrams and cell population data generated by fully automated hematology analyzers: New tools for screening malaria infection? *Int J Lab Hematol* 2018;40:326-34.
30. Lippi G, Plebani M. Capillary electrophoresis for the screening and diagnosis of inherited hemoglobin disorders.

- Ready for prime time? *Clin Chem Lab Med* 2016;54:5-6.
31. Wild BJ, Green BN, Cooper EK, et al. Rapid identification of hemoglobin variants by electrospray ionization mass spectrometry. *Blood Cells Mol Dis* 2001;27:691-704.
 32. Harteveld CL. State of the art and new developments in molecular diagnostics for hemoglobinopathies in multiethnic societies. *Int J Lab Hematol* 2014;36:1-12.
 33. Traeger-Synodinos J, Harteveld CL, Old JM, et al. EMQN Best Practice Guidelines for molecular and haematology methods for carrier identification and prenatal diagnosis of the haemoglobinopathies. *Eur J Hum Genet* 2015;23:560.
 34. Steensma DP. New challenges in evaluating anemia in older persons in the era of molecular testing. *Hematology Am Soc Hematol Educ Program* 2016;2016:67-73.
 35. Lippi G, Bassi A, Bovo C. The future of laboratory medicine in the era of precision medicine. *J Lab Precis Med* 2016;1:7.
 36. Nasybullina EI, Nikitaev VG, Pronichev AN, et al. Expert diagnostic system for hemoglobinopathies using the data on blood, erythrocyte, and hemoglobin state. *Bull Lebedev Phys Inst* 2015;42:206.
 37. Amendolia SR, Brunetti A, Carta P, et al. A real-time classification system of thalassemic pathologies based on artificial neural networks. *Med Decis Making* 2002;22:18-26.

doi: 10.21037/jlpm.2018.07.09

Cite this article as: Lippi G, Plebani M. Recent developments and innovations in red blood cells diagnostics. *J Lab Precis Med* 2018;3:68.